SYNTHESIS OF [²H]₅-EPITESTOSTERONE

Kristiina Wähälä,*1 Tiina Väänänen,1 Tapio Hase1 and Antti Leinonen2

¹Department of Chemistry, P.O. Box 55, FIN-00014 University of Helsinki, Finland Internet: Kristiina.Wahala@helsinki.fi. Fax 358-0-191 403 66.
²United Laboratories Ltd., P.O. Box 222, FIN-00381 Helsinki, Finland.

SUMMARY

Isotopically pure d_5 -epitestosterone has been synthesized for the first time. This polydeuterated 17 α -hydroxyandrosterone was prepared by H/D exchange reaction with MeOD-D₂O under basic conditions. Its characteristics are described in detail, including its enolization into a d_4 -heterodienic silyl ether derivative upon silylation.

Keywords: [²H]₅-17α-Hydroxyandrost-4-en-3-one, d₅-epitestosterone, deuteration, labeling, synthesis, dienolsilyl ether

INTRODUCTION

Deuterated epitestosterone (17 α -hydroxyandrost-4-en-3-one, 17 α -cis-testosterone) was required as a standard for the determination of urinary epitestosterone in doping analysis. The quantitation is performed by gas chromatography - mass spectrometry selected ion monitoring (GC/MS SIM) using the standard screening procedure for anabolic steroids.¹ In this procedure, the urine sample is extracted using either XAD-2 resin or bonded phase column (C18) followed by enzymatic hydrolysis of steroid glucuronides, extraction with diethyl ether and derivatization of steroids to trimethylsilyl ethers.

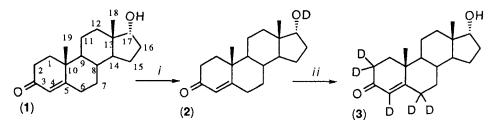
For quantitation, the use of a polydeuterated analog as an internal standard is highly recommended. It is desirable that the internal standard is labelled by at least 3-4 deuterium atoms in order to obtain a straight line calibration curve free of interference from the compound to be measured. In addition, the deuterated internal standard should be compatible with all sample preparation steps because any re-exchanges of C—D to C—H can not be tolerated.

 d_1 - And d_2 - labeled derivatives of epitestosterone have been synthesized previously from 3-methoxy-3,5-androstadiene-17-one.² The latter was reduced by NaBD₄ to give a mixture of 17- d_1 -testosterone and 17- d_1 -epitestosterone in an unspecified yield and ratio in about 90% isotopic purity, the remainder consisting of unlabeled products. 16,16- d_2 -Epitestosterone was prepared by deuteration of 3-methoxy-3,5-androstadiene-17-one with NaOD/CD₃OD followed by NaBH₄ reduction, hydrolysis and HPLC purification to afford a 0.7% yield of 16,16- d_2 -epitestosterone.² It is not clear why the expensive CD₃OD had to be used here instead of CH₃OD as the methanol methyl group is not involved in any reactions.

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RESULTS AND DISCUSSION

Since we required epitestosterone with a minimum of three D atoms we envisaged a Mitsunobu inversion³ of the commercially available 16,16,17- d_3 -testosterone. Unexpectedly, this reaction was unsuccessful and we will report a study of this failure separately. However, a base catalyzed H/D exchange reaction⁴ of 17-O-d-epitestosterone (2) in MeOD-D₂O furnished 2,2,4,6,6-[²H]₅-epitestosterone (3) in quantitative yield and with a d_5 isotopic purity over 90% according to ¹³C-NMR and MS data. After the exchange reaction the product was treated with a large excess of H₂O to reinstate the protic hydroxy group so as to avoid ambiguities resulting from uncontrolled OD/OH exchange.



Scheme 1. Synthesis of d_5 -epitestosterone (3) from predeuterated epitestosterone (2). $i : D_2O$, ii MeONa, MeOD, D_2O , then H_2O .

As expected, the d_5 -deuterated epitestosterone (3) performs very satisfactorily in GC/MS SIM analysis, despite the fact that one D atom is lost when the standard is trimethylsilylated using trimethyliodosilane⁵ as a catalyst. This is due to enolization⁵ of the 3-keto group to give, after silylation, the heterodienol silyl ether (4) in 99% yield and pure by GC. This enol ether (4) produces an abundant molecular ion (M⁺ 436, 100%) in the mass spectrum increasing the sensitivity for the selected ion monitoring (SIM) quantitation. The structure of this dienol silyl ether was established by NMR, UV and MS spectroscopy, all the spectral data being completely incompatible with the alternative homodienic silyl ether structure. To our knowledge there is no literature report of the exact structure of the dienol silyl ether of testosterone or epitestosterone although the enolization is known to occur on silylation with a number of catalysts.

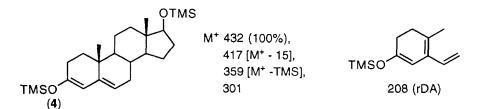


Figure 1. The dienol silyl ether of epitestosterone (4) and its main fragments in the mass spectrum.

EXPERIMENTAL

The m.p. was determined in an open capillary tube with an Electrothermal apparatus, and is uncorrected. ¹H and ¹³C NMR spectra were recorded on a Varian GEMINI-200 FT NMR spectrometer for solutions in CDCl₃ and CD₃COCD₃ (Me₄Si as the internal standard). Mass spectra were obtained with a Jeol JMS SX102 mass spectrometer operating at 70 eV, or 10-20 eV when determining the deuterium content. Samples were introduced at 120-150°C by a direct-inlect probe. TLC was conducted on Merck silica gel 60 F₂₅₄ plates with CH₂Cl₂-EtOAc (7:2) elution. *d₅*-Epitestosterone was fully characterized by ¹H, ¹³C NMR and DEPT spectra, LRMS and HRMS analysis and the compound was homogenous by TLC. In the ¹³C NMR spectrum the deuterated methine carbon appears as a low-intensity triplet (t) in the proton noise decoupled spectrum as compared to the singlet in the spectrum of undeuterated material.

2,2,4,6,6-[²H]₅-epitestosterone

Epitestosterone (1) was suspended in D₂O and the heavy water was evaporated by rotary evaporator to give the predeuterated epitestosterone (2). Slices of clean solid sodium (0.2 g, 8.7 mmol) were added slowly into MeOD (10 ml) under argon followed by 2 (0.1 g, 0.35 mmol). The yellowish reaction mixture was heated to reflux, D₂O (1.5 ml) added and the refluxing continued for 1.5 h. After the reaction MeOD and D₂O were evaporated off

. The residue was dissolved in D₂O and acidified with 2 N H₂SO₄. The precipitated crude product (100 %) was collected by filtration and recrystallized from methanol, m.p. 210-212 °C. In ¹H-NMR the undeuterated signals at δ 5.73 (H-4, 1H), 2.15 (H-2, 2H) are missing; ¹³C- NMR δ 17.3 (C-18), 17.8 (C-19), 20.9 (C-11), 24.9 (C-15), 31.5 (C-16), 32.3 (C-7), 32.5 (C-6)^t, 32.7 (C-2)^t, 35.9 (C-1 & C-8), 36.1 (C-12), 38.9 (C-10), 45.5 (C-13), 48.5 (C-14), 54.0 (C-9), 80.0 (C-17), 129.6 (C-4)^t, 171.8 (C-5), 200.3 (C-3); *m/z* 293 (M⁺, 46%), 291 (35), 276 (32), 260 (14), 247 (19), 231 (19), 147 (65), 129 (41), 105 (17), 93 (27) (Found: M+ 293.2408. C₁₉H₂₃O₂D5 requires 293.2413).

Silylation

Epitestosterone was derivatized by 50µl of a solution of *N*-Methyl-*N*-trimethylsilyltrifluoroacetamide, trimethyliodosilane and dithioerythritol (1000:2:4), at 60 °C for 15 min yielding 99 % of the corresponding dienol silyl ether (4), λ_{max} (hexane)/nm 242, 290; ¹H-NMR δ 5.27 (H-4, 1H), 5.16 (H-6, 1H), 3.63 (H-17, 3-H); ¹³C- NMR δ 151.2 (C-3), 143.5 (C-5), 120.9 (C-6), 111.4 (C-4), 84.3 (C-17), 53.8 (C-9), 51.3 (C-14), 45.8 (C-13), 39.6 (C-2), 36.7 (C-12), 34.7 (C-8), 34.0 (C-1), 33.5 (C-7), 32.9 (C-10), 30.3 (C-16), 24.0 (C-15), 23.5 (C-11), 21.2 (C-19), 20.7 (C-18).

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REFERENCES

- Donike M., Geyer H., Gotzmann A., Kraft M., Mandel F., Nolteernsting E., Opfermann G., Sigmund G., Schänzer W. and Zimmermann J.— 1988 Dope Analysis. In: Bellotti P., Benzi G., Ljungqvist A., eds.: Official Proceedings of International Athletic Foundation World Symposium on Doping in Sport - 1987, 53-80, London, (1988). Published by the International Athletic Foundation.
- Wood A.W., Swinney D.C., Thomas P.E., Ryan D.E., Hall P.F., Levin W. and Garland W.A. — J. Biol. Chem. 263: 17322 (1988).
- 3. Martin S.F. and Dodge J.A. -- Tetrahedron Lett. 32: 3017 (1991).
- Shapiro R.H, Williams D.H., Budzikiewicz H. and Djerassi C. J. Am. Chem. Soc. 86: 2837 (1964).
- 5. Donike M. and Zimmerman J. J. Chromatogr. 202: 483 (1980).